Ethyl 1-(2-Methylbutyl)-2-oxocyclohexane-1-carboxylate (58). Treatment of 0.030 g of 20 with excess of active Raney nickel in 5 mL of refluxing ethanol for 6 h followed by filtration and evaporation of the ethanol left 0.015 g of oily 58: 63% yield; IR 1740, 1720 cm⁻¹; NMR δ 0.9 (t, 3), 1.1–2.5 (m, 18, 3.0–3.4 (m, 3), 4.1 (dq, 2); mass spectrum, m/e 242 (M⁺), 170, 124, 95, 81; HRMS, m/e 242.188 (C₁₄H₂₆O₃ requires 242.188).

An authentic sample of 58 was prepared by generation of the sodium enolate of 19 by using NaH in THF and treatment with 2-methyl-1-bromobutane. This product was identical in all respects with 58.

2-(2-Methylbutyl)cyclohexanone (56). A solution of 3.51 g (2×10^{-2} mol) of 1-pyrrolidinylcyclohexene with 3.5 g (2×10^{-2} mol) of 1-bromo-2-methylbutane in 20 mL of dioxane was heated at reflux for 20 h. The mixture was diluted with 10 mL of 3 N HCl and stirred for 1 h. The usual workup afforded 1.95 g of oil from which an analytical sample of 56 was obtained by preparative GC (column C): IR 1710 cm⁻¹; NMR δ 1.0 (d, $J \approx 8$ Hz, 3), 1.1 (d, $J \approx 8$ Hz, 3), 1.4–2.5 (m, 14); mass spectrum, m/z (relative intensity) 168 (M⁺, 5), 98 (base, 100).

A solution of 0.050 g of 5 in 5 mL of ethanol was treated with about 0.1 g of activated Raney nickel and heated at reflux for 3 h. Filtration and evaporation left 0.035 g of a mixture containing ca. 45% of 56, identical with the 56 above by GC (columns A, C, D). None of the remaining compounds in this mixture were 57 (GC/MS).

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Registry No. 1, 70109-88-5; 2, 39198-55-5; 3, 108-94-1; 4, 70234-67-2; 5, 83720-11-0; 6, 6651-36-1; 7, 13482-23-0; 8, 83720-12-1; 9, 83720-13-2; 10, 98-53-3; 11, 83720-14-3; 12, 83720-15-4; 13, 19980-35-9; 14, 83720-16-5; 15, 583-60-8; 16, 19980-33-7; 17, 83720-17-6; 18, 83720-18-7; 19, 1655-07-8; 20, 83720-19-8; 21, 83720-21-2; 22, 83720-22-3; 23, 502-42-1; 24, 70109-90-9; 25, 83720-23-4; 26, 502-49-8; 27, 83720-24-5; 28, 83720-25-6; 29, 83720-26-7; 30, 57044-58-3; 31, 83780-83-0; 32, 83720-27-8; 33, 83720-28-9; 34, 70109-89-6; 35, 83780-28-3; 36, 818-23-5; 37, 53282-55-6; 38, 83720-29-0; 39, 83720-30-3; 40, 123-19-3; 41, 83720-31-4; 42, 57641-21-1; 43, 83720-32-5; 44, 609-14-3; 45, 83720-33-6; 46, 83720-35-8; 47, 83720-36-9; 48, 83720-37-0; 49, 83720-38-1; 50, 83720-40-5; 51, 83720-41-6; 52, 33066-07-8; 53, 1654-87-1; 55, 3574-58-1; 56, 20118-23-4; 58, 83720-42-7; 59, 78945-46-7; 1,4-dichloro-1,3-butadiene, 2984-42-1; dimethyl sulfide, 75-18-3; 1-pyrrolidinylcyclohexene, 1125-99-1; 1-bromo-2methylbutane, 10422-35-2.

Natural Ferric Ionophores: Total Synthesis of Schizokinen, Schizokinen A, and Arthrobactin

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The synthesis of the microbial iron chelators schizokinen (1) and arthrobactin (2) are described. O-Benzyl-N-(carbobenzyloxy)hydroxylamine (7) was subjected to triphenylphosphine/diethyl azodicarboxylate mediated alkylation with alcohol amine 10 to give the N-alkylated product 13, which was converted to the protected 1-amino-3-(acetylhydroxyamino)propane 4a by hydrogenation in the presence of acetic anhydride. Several citric acid derivatives were prepared which were activated at both terminal carboxyl groups and protected at the internal carboxyl groups. 1-Amino-3-[(benzyloxy)amino]propane p-toluenesulfonic acid double salt (19) was coupled to citric acid derivative 28c to give protected schizokinen 29c, which was deprotected in two steps to yield schizokinen (1). The 1-amino-5-(acetylhydroxyamino)pentane derivative 4b was deprotected and coupled with citric acid derivative 28a to give 31, which was deprotected in two steps to yield arthrobactin (2). Preliminary attempts to synthesize schizokinen resulted in formation of succinimide 26. Reductive debenzylation of 26 provided 33 which was shown to be identical with schizokinen A.

Iron is an essential element for all life forms. Although iron is one of the most abundant elements, the extreme insolubility of ferric ion at neutral and alkaline pH places severe restrictions on its metabolism. Iron absorption from the diet is physiologically controlled, but the body has no regulatory mechanism for eliminating a toxic excess introduced by accidental overdose or by multiple transfusions. Cooley's anemia and its transfusional treatment provide an example of the difficulty of correcting deficient iron metabolism. According to the World Health Organization, the group of diseases called the thalassemias, of which Cooley's anemia is the most severe, is the largest health problem in the world for single-locus genetic diseases. Extensive iron overload induced by the multiple transfusions during treatment of Cooley's anemia causes deposition of the metal in the heart, liver, endocrine glands,

In principle, iron overload can be treated by administration of an iron-chelating agent to promote remobilization and excretion of the deposited iron. Perhaps the best models for iron chelation are provided by microbial systems which have envolved highly specific and efficient iron-sequenstering agents.² These siderophores primarily utilize either hydroxamic acids³ or catechols⁴ for the che-

and other organs. The ultimate result is organ malfunction and early death. $^{\rm 1}$

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Natural Ferric Ionophores

lating ligands. Most of the hydroxamate-containing siderophores, including ferrichrome,⁵ rhodotorulic acid,⁶ coprogen,⁷ aerobactin,⁸ and mycobactins,⁹ contain ω -Nhydroxy L-amino acids. However, others, such as schizokinen (1),¹⁰ arthrobactin (2),¹¹ and ferrioxamine B,¹² utilize 1-amino- ω -(hydroxyamino)alkane residues to bind ferric ion. During evolution, these residues may have resulted from decarboxylation¹³ of the corresponding ω -Nhydroxy α -amino acids.

Subsequent to completion of the synthesis of aerobactin,¹⁴ schizokinen (1) and arthrobactin (2) have been among

$$\begin{array}{cccc} \mathsf{CH}_3 & \mathsf{CH}_3 & \mathsf{CH}_3 & \mathsf{CH}_3 \\ \mathsf{C}{:}\mathsf{O} & \mathsf{C}{:}\mathsf{O} & \mathsf{C}{:}\mathsf{O} & \mathsf{C}{:}\mathsf{O} \\ \mathsf{N}{-}\mathsf{O}\mathsf{H} & \mathsf{N}{-}\mathsf{O}\mathsf{H} & \mathsf{N}{-}\mathsf{O}\mathsf{H} \\ (\mathsf{CH}_2)_n & \mathsf{COOH} & (\mathsf{CH}_2)_n & (\mathsf{CH}_2)_n \\ \mathsf{N}\mathsf{H}{-}\mathsf{CO}{-}\mathsf{CH}_2{-}\mathsf{C}{-}\mathsf{C}\mathsf{H}_2{-}\mathsf{CO}{-}\mathsf{N}\mathsf{H} & \mathsf{N}\mathsf{H}_2 \\ & \mathsf{O}\mathsf{H} \\ & \mathsf{1} & (\mathsf{n}{:}3) & \mathsf{3} & \mathsf{a} (\mathsf{n}{:}3) \\ \mathsf{2} & (\mathsf{n}{:}5) & \mathsf{b} (\mathsf{n}{:}5) \end{array}$$

the targets of recent synthetic studies in our laboratory. These compounds are growth factors for Bacillus megaterium and Arthrobacter pascens, respectively. Schizokinen consists of two residues of 1-amino-3-(N-acetylhydroxyamino)propane (3a) linked to the two terminal carboxyl groups of a citric acid residue by amide bonds.¹⁰ Similarly, arthrobactin is composed of two 1-amino-5-(Nacetylhydroxyamino)pentane (3b) residues linked to citric acid. The 1-amino-n-(N-acetylhydroxyamino)alkane fragments were initially considered to constitute the main challenge in the synthesis of 1 and 2. Conceptually, the suitably protected forms 4a and 4b can be derived from the alkylation of O-protected hydroxamates with derivatives of commercially available amino alcohols 5a,b (eq 1).



However, alkylation of O-substituted hydroxamates may occur on the carbonyl oxygen as well as on the nitrogen.¹⁵ Thus, control over this feature was most important for our synthetic purpose, and a minimum requirement of predominant N-alkylation was essential. Consequently, studies related to the alkylation of hydroxamates were



performed by varying the leaving group on the alkylating agent, the base which generated the hydroxamate anion. and the acyl group of the hydroxamate.

In addition to the hydroxyamino fragments, a citric acid component was required which was activated at both terminal carboxyl groups and protected at the internal carboxyl and hydroxyl groups. Coupling of such a synthon with 4a or 4b followed by complete deprotection was expected to provide schizokinen (1) and arthrobactin (2).

Results and Discussion

Preparation of the Hydroxylamine Compounds. As in the synthesis of aerobactin,¹⁴ the benzyl group was chosen as the hydroxamate O-substituent because of its stability toward planned reactions and yet its ease of eventual removal. The required primary O-benzyl hydroxamates were prepared by the direct acylation of Obenzylhydroxylamine (OBHA, Scheme I). In this manner, O-benzyl acetohydroxamate (6) and O-benzyl-N-(carbobenzyloxy)hydroxylamine (7) have been previously prepared.^{14,16} O-Benzyl-N-(tert-butoxycarbonyl)hydroxylamine (8) was prepared by the acylation of OBHA with di-tert-butyl dicarbonate in THF. O-Benzyl-N-[p-(nitrophenyl)carbobenzoxy]hydroxylamine (9) was prepared by acylation of OBHA with *p*-nitrobenzyl chloroformate.

Hydroxamate alkylations were performed by two methods; treatment of the corresponding hydroxamate anion with an alkyl halide and direct alkylation of hydroxamates with alcohols by using azodicarboxylates and triphenylphosphine (TPP).¹⁴ In the first approach (Scheme II), the amino group of 5 was protected with the tert-butoxycarbonyl (Boc) group to provide 10. The alcohol 10 was then converted to the bromide 11 with TPP and CBr₄. Treatment of 11 with O-benzyl acetohydroxamate (6) and K_2CO_3 in acetone containing a catalytic amount of KI¹⁴ provided alkylated hydroxamates in 75-85% yield. However, a 4:1 ratio of N- to O-alkylated products (4 and 12) was obtained. Isomers 4b and 12b were easily separated by chromatography, but 4a and 12a

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were not. After the initial chromatography of 4a and 12a, fractions were obtained which were isographic on TLC (R_{i}) 0.24; hexanes/ethyl acetate, 65:35), but the ^{1}H NMR spectrum clearly indicated the presence of both 4a and 12a. Repetitive chromatography eventually provided 4a in 40-50% yield. Several different base and solvent system combinations were subsequently tried for the alkylation of 11a, but none improved the N- to O-alkylation ratio.

Attempted alkylation of O-benzyl acetohydroxamate (6) with 10 in the presence of DEAD/TPP gave predominant O-alkylation. However, the DEAD/TPP-mediated alkylation of the O-benzyl-N-(alkoxycarbonyl)hydroxylamines 7 and 9 with 10a (Scheme III) gave the desired N-alkylated products 13a and 13b in 70–90% yields with no competitive O-alkylation. Careful catalytic hydrogenation of 13a with 5% Pd/C selectively removed the carbobenzyloxy (Cbz) group. When the reaction was performed in the presence of acetic anhydride, the desired N-acetylated product 4a was obtained in 65% yield. Longer hydrogenation removed both benzyl groups to provide the diacetate 14. Methanolysis of 14 with a catalytic amount of NH_3 gave the hydroxamic acid 15 in quantitative yield. The same sequence was attempted by starting with the *p*-nitrobenzyl carbamate 13b, but, surprisingly, less selective hydrogenation was observed.

Mild acid treatment was anticipated to remove the Boc protecting group from 4a and 4b to provide the free amines 16a and 16b, respectively. Indeed, treatment of 4b with trifluoroacetic acid (TFA) gave 16b cleanly (eq 2). How-

$$\begin{array}{c} \mathsf{BOC-NH} - (\mathsf{CH}_2)_5 \cdot \mathsf{N} - \mathsf{C}^{-}\mathsf{CH}_3 & \xrightarrow{1. \text{ TFA}} & \mathsf{NH}_2 - (\mathsf{CH}_2)_5 \cdot \mathsf{N} - \mathsf{C}^{-}\mathsf{CH}_3 & (2) \\ \circ \mathsf{CH}_2\mathsf{Ph} & \xrightarrow{2. \text{ base}} & \mathsf{NH}_2 - (\mathsf{CH}_2)_5 \cdot \mathsf{N} - \mathsf{C}^{-}\mathsf{CH}_3 & (2) \\ \circ \mathsf{CH}_2\mathsf{Ph} & \circ \mathsf{CH}_2\mathsf{Ph} & \circ \mathsf{CH}_2\mathsf{Ph} \end{array}$$

ever, the reaction of 4a with either TFA or p-toluene-



sulfonic acid (TsOH) followed by the usual workup gave the rearranged product 18 which apparently resulted from acyl transfer (Scheme IV). Further treatment of 18 with TsOH (or TsOH \cdot H₂O) gave the disalt 19. Compound 19 could be obtained directly from 4a or the diBoc-protected analogue 21 (Scheme V) with excess TsOH·H₂O. Compound 21 was obtained in 41% yield by DIPAD (diisopropyl azodicarboxylate)/TPP-mediated reaction of alcohol 10a with the O-benzyl-N-(tert-butoxycarbonyl)hydroxylamine (8). Finally, in contrast to the reactions of 4a with TFA or TsOH, treatment of 4a with anhydrous HCl gave 16a (X = Cl) in 97% yield.

Citric Acid Derivatives and the Coupling Reactions. Only two of the carboxyl groups of citric acid 22 are involved in the amide linkages of schizokinen and arthrobactin. Consequently, the remaining carboxylic acid and hydroxyl groups must be protected during the synthesis. Anhydromethylenecitric acid (23), readily prepared from citric acid and paraformaldehyde,¹⁸ was considered an appropriate starting material. The corresponding diacid chloride 24 was obtained by treatment of 23 with PCl₅¹⁷ (Scheme VI). Reaction of 24 with the amine 16a (200 mol %, Et₃N, 0 °C, 2 h) provided two major products as indicated by TLC analysis. Spectral analysis after the workup indicated that one component was the desired material 25, and the other appeared to be the imide 26. Presumably 26 results from intramolecular opening of the anhydromethylene unit of 25. Facile imide formation has been observed in several similar systems.^{18,19,20,22} Attempted chromatographic separation of 25 and 26 on silica

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Scheme VII



gel resulted in further conversion of 25 to 26. In another control, treatment of 25 and 26 with Et_3N for 1 h at room temperature resulted in complete conversion of 25 to 26. Interestingly, during the original isolation of schizokinen, another unidentified ferric chloride positive component, schizokinen A, was obtained.¹⁰ Schizokinen A reportedly could also be prepared from schizokinen by heating. Our observation of the ease of formation of the imide 26, coupled with comparison of the chemical and spectral properties of 26 and those reported for schizokinen A, prompted us to suggest the imide structure (debenzylated 26) for schizokinen A. Further comparison of the properties of debenzylated material 33 (Scheme VIII) with data provided by Professor Nielands^{10b,c} confirmed 33 as the structure for schizokinen A.

In order to avoid formation of imide 26, we considered other protected citric acid derivatives. Thus, reaction of anhydromethylenecitric acid (23) with a variety of alcohols in the presence of excess Et₃N produced the monoesters 27 (Scheme VII). The bis(p-nitrophenyl) esters 28 were then prepared by reaction of 27 with dicyclohexylcarbodiimide (DCC) and p-nitrophenol. The reaction of 28a and the amine 16a in the presence of a number of tertiary bases gave three products and recovered ester 28. The desired product 29 and the monoamide 30 were obtained in only $\sim 10\%$ yield. The major product was 18 which again appeared to result from intramolecular acyl transfer of 16a. However, no imides were obtained nor were products observed which would result from attack of the hydroxylamine portion of 18 on the mono- or bis(p-nitrophenyl) esters 30 and 28, respectively.

This latter observation prompted us to attempt the coupling of 28 with the previously prepared 1-amino-3-[(benzyloxy)amino]propane 19. Indeed, the reaction of 28a with 19a followed by acetylation with acetic anhydride did provide the desired bis amide 29a (eq 3). Spectral analysis

$$28 + 29 \rightarrow \xrightarrow{(CH_3CO)_2O} 26 + 29 \tag{3}$$

of the crude reaction mixture also indicated the succinimide 26. Attempted chromatographic purification of 29 again resulted in its conversion of 26. In fact, just dissolving the mixture in polar protic solvents like 2-propanol promoted imide formation. Similar results were obtained upon attempting the coupling reaction with the methyl ester 28b.

In order to minimize the imide formation, we decided to attempt the coupling reaction of 19 with 28c, the bis-

Scheme VIII



(*p*-nitrophenyl) ester containing the more hindered isopropyl group to protect the internal carboxyl group of the citric acid. The reaction was performed as before with subsequent addition of acetic anhydride to acetylate the hydroxylamine groups before the workup. In this case the desired product **29c** was obtained in 75.6% yield with no contamination by the imide **26**.

In contrast to the problems associated with the coupling of 28a and 19, the reaction of 28a with 16b [1-amino-5-[(benzyloxy)acetylamino]pentane] provided the desired protected arthrobactin 31 cleanly (eq 4). In this case the



free amine of 16b could be used directly since it is not susceptible to the intramolecular acyl-transfer reaction observed with 16a. Thus, the absence of added base to liberate the free amine appears to help avoid undesired imide formation. The longer hydrocarbon chain, or branching in the chain as in our aerobactin synthesis,¹⁴ also seems to diminish competitive imide formation.

All that remained for the synthesis of schizokinen (1) and arthrobacin (2) was respective deprotection of 29 and 31. Initial attempts at the catalytic hydrogenation of 29a indicated that polar solvents were required to avoid precipitation of the product on the catalyst. However, when 29a was hydrogenolyzed in methanol, the monomethyl ester 32 was obtained (Scheme VIII). This again indicated the lability of the internal protected carboxyl group. A change of the solvent to THF-H₂O resulted in cyclization and eventual production of the imide 33 (schizokinen A) as the major product and very little of the desired schizokinen (1). Similarly direct hydrogenation of 31 did not produce arthrobactin (2) cleanly. Alternatively, 29c and 31 were separately saponified with aqueous NaOH in THF. The resulting sodium carboxylates 34a and 34b were debenzylated with H_2 and Pd/C to provide schizokinen (1) and arthrobactin (2) cleanly (Scheme IX). Thus, the carboxylate anions of **34a** and **34b** effectively prohibits the undesired imide formation.

In conclusion, the alkylation of O-substituted hydroxamates offers an attractive route to the constituents of the natural hydroxamate siderophores. The versatility of the resulting intermediates has been demonstrated by the total synthesis of the natural microbial iron chelators schizokinen and arthrobactin. The chemistry involved has also led to the determination of the structore of schizokinen A. Further studies are being directed toward the synthesis of analogues of the microbial iron chelators which may be clinically useful for teatment of iron overload.

Experimental Section

General Methods. Melting points were taken on a Thomas-Hoover capilary melting point apparatus and are uncorrected. Infrared spectra (IR) were recorded on a Perkin-Elmer 727B spectrophotometer. Proton NMR spectra were obtained on a Varian A60, EM-390 or XL-100 spectrometer in deuteriochloroform (unless otherwise stated) and are reported in parts per million downfield of internal tetramethylsilane (δ units).

Mass spectra were recorded on an AEI Scientific Apparatus MS902 or Du Pont DP 102 spectrometer. Elemental analyses were performed by Midwest Microlabs. Field desorption mass spectra were obtained by Mr. John L. Occolwitz (Eli Lilly and Co.).

O-Benzyl Acetohydroxamate (6). This was prepared by the method of Nicolaus et al.¹⁶ except that the product was purified by acid/base extraction rather than by vacuum distillation.

O-Benzyl-N-(carbobenzyloxy)hydroxylamine (7). O-Benzylhydroxylamine hydrochloride (3.19 g, 0.02 mol) was suspended in dry acetonitrile (40 mL) and treated with pyridine (3.23 mL, 0.04 mol). After the mixture was cooled to 0 °C, benzyl chloroformate (2.85 mL, 0.02 mol) was added dropwise with stirring. The mixture was allowed to warm to room temperature and vigorously stirred for 24 h. Volatile components were evaporated, and the residue was taken up in ethyl acetate. This was washed twice with 0.6 N HCl, once with H₂O, once with 0.6 M NaHCO₃, and once again with H₂O. After the mixture was dried (MgSO₄) and the solvent evaporated, the residue was crystallized from ethyl acetate/hexane to yield 7: 3.09 g (60%); colorless crystals; mp 66-67 °C (lit.²¹ mp 65-68 °C); ¹H NMR (CDCl₃) δ 4.8 (s, 2 H), 5.12 (s, 2 H), 7.32 (s, 10 H), 7.63 (s, 1 H, NH).

O-Benzyl-N-(*tert***-butoxycarbonyl)hydroxylamine** (8). O-Benzylhydroxylamine hydrochloride (6.38 g, 0.04 mol) was suspended in THF/H₂O (1:1, 80 mL) and treated with NEt₃ (triethylamine; 5.49 mL, 0.04 mol). Di-*tert*-butyl dicarbonate (8.72 g, 0.04 mol) dissolved in THF (20 mL) was added dropwise with stirring for 30 min at room temperature. After the addition was completed, the reaction mixture was allowed to stir 1.5 h more at room temperature. Volatile components were evaporated, and the residue was taken up in ethyl acetate. This was washed twice with 0.5 M citric acid and once with H₂O. After the mixture was dried (MgSO₄) and the solvent evaporated, the residue was crystallized from hexane to yield 8: 8.20 g (92%); colorless crystals; mp 45–47 °C; ¹H NMR (CDCl₃) δ 1.45 (s, 9 H), 4.84 (s, 2 H), 7.38 (s, 5 H), 7.75 (s, 1 H, NH). Anal. Calcd for C₁₂H₁₇NO₃: C, 64.55; H, 7.67; N, 6.27. Found: C, 64.64; H, 7.70; N, 6.25.

O-Benzyl-N-(p-nitrobenzyloxycarbonyl)hydroxylamine (9). O-Benzylhydroxylamine hydrochloride (3.19 g, 0.02 mol) was suspended in dry acetonitrile (40 mL) and treated with pyridine (3.23 mL, 0.04 mol). p-Nitrobenzyl chloroformate (4.31 g, 0.02 mol) dissolved in acetonitrile (20 mL) was added dropwise with stirring for 30 min at room temperature. After the addition was completed, the reaction mixture was allowed to stir for 24 h. Volatile components were evaporated, and the residue was taken into ethyl acetate. This was washed twice with 0.6 N HCl once with H₂O, once with 0.6 M NaHCO₃, and once again with H₂O. After the mixture was dried and the solvent evaporated, the residue was crystallized from ethyl acetate/hexane to yield 9: 3.42 g (57%); light yellow crystals; mp 96–97 °C; ¹H NMR (CDCl₃) δ 4.87 (s, 2 H), 5.23 (s, 2 H), 7.40 (s, 5 H), 7.67 (d, 2 H), 8.20 (d, 2 H). **3-[(tert-Butoxycarbonyl)amino]-1-propanol** (10a). 3-Amino-1-propanol (7.5 g, 0.1 mol; Aldrich) was dissolved in THF/H₂O (1:1, 150 mL). Di-tert-butyl dicarbonate (21.8 g, 0.1 mol; Sigma) dissolved in THF (50 mL) was added dropwise with stirring for 30 min at room temperature. After the addition was completed, the reaction mixture was allowed to stir 2 h more at room temperature. Volatile components were evaporated, and the residue was taken up in ethyl acetate. This was washed twice with 0.5 M citric acid and once with H₂O. After the mixture was dried (MgSO₄) and the solvent evaporated, the residue was an oil: 12.6 g (72%); ¹H NMR (CDCl₃) δ 1.43 (s, 9 H), 1.5–1.9 (m, 2 H), 3.23 (q, 2 H), 3.63 (t, 2 H), 4.17 (s, 1 H, OH), 5.70 (s, 1 H, NH).

5-[(tert-Butoxycarbonyl)amino]-1-pentanol (10b). 5-Amino-1-pentanol (10.31 g, 0.1 mol; Aldrich) was dissolved in acetonitrile (200 mL). Di-tert-butyl dicarbonate (21.8 g, 0.1 mol) dissolved in acetonitrile (50 mL) was added dropwise with stirring for 30 min and the mixture allowed to stir 24 h at room temperature. Volatile components were evaporated, and the residue was taken up in ethyl acetate. This was washed twice with 0.5 M citric acid and once with H₂O. After the mixture was dried (MgSO₄) and the solvent evaporated, the residue was vacuum desiccated overnight to yield an oil: 18.70 g (92%); ¹H NMR (CDCl₃) δ 1.2-1.7 (m, 15 H), 3.08 (q, 2 H), 3.58 (t, 2 H), 3.97 (s, 1 H, OH), 5.42 (s, 1 H, NH).

3-[(tert-Butoxycarbonyl)amino]-1-propyl Bromide (11a). 3-[(tert-Butoxycarbonyl)amino]-1-propanol (10a; 6.4 g, 0.0365 mol) and PPh₃ (12.45 g, 0.0485 mol) were dissolved in dry THF (150 mL) and treated dropwise with CBr₄ (15.75 g, 0.0475 mol) in acetonitrile (60 mL) such that the temperature did not rise much above ambient. After the mixture was stirred at room temperature for 6 h, the solvent was evaporated, and the residue was chromatographed on silica gel (4 × 60 cm), eluting with ethyl acetate/hexane (20:80). 11a was crystallized from hexane: 6.52 g (75%); mp 38-39 °C; ¹H NMR (CDCl₃) δ 1.43 (s, 9 H), 1.8-2.2 (m, 2 H), 3.1-3.4 (m, 4 H), 5.10 (s, 1 H, NH). Anal. Calcd for C₈H₁₆NO₂Br: C, 40.35; H, 6.77; N, 5.88. Found: C, 40.17; H, 7.01; N, 5.89.

5-[(tert-Butoxycarbonyl)amino]-1-pentyl Bromide (11b). 5-[(tert-Butoxycarbonyl)amino]-1-pentanol (10b; 2.03 g, 10 mmol) and PPh₃ (3 g, 11.5 mmol) were dissolved in dry THF (50 mL). CBr₄ (3.8 g, 11.5 mmol) in THF (15 mL) was added dropwise over 2 h at room temperature. The reaction was allowed to stir overnight. The solvent was evaporated, and the residue was chromatographed on silica gel (2 × 60 cm), eluting with ethyl acetate/hexanes (20:80). Compound 11b was crystallized from hexane: 2.08 g (78%); mp 26–28 °C; ¹H NMR δ 1.2–1.7 (m, 13 H), 1.7–2.0 (m, 2 H), 3.15 (q, 2 H), 3.40 (t, 2 H), 4.80 (br s, 1 H, NH); mass spectrum (CI with Ar), m/e 210 (M – 55, for HO₂C– NH₂⁺-(CH₂)₅–Br).

1-[(tert-Butoxycarbonyl)amino]-3-[acetyl(benzyloxy)amino]propane (4a). 3-[(tert-Butoxycarbonyl)amino]-1-propyl bromide (11a; 1.79 g, 7.5 mmol) O-benzyl acetohydroxamate (6; 2.5 g, 15 mmol), KI (0.2 g, 1.2 mmol), and anhydrous K₂CO₃ (5.52 g, 50 mmol) were placed in dry acetone (30 mL) and refluxed for 24 h. After filtration and evaporation, the residue was taken up in ether. This was washed twice with 0.5 N NaOH and once with H_2O . After the mixture was dried and the solvent evaporated, the residue was chromatographed on silica gel $(2 \times 50 \text{ cm})$, eluting with $\text{CH}_2\text{Cl}_2/i\text{-}\text{PrOH}$ (98:2) several times. The product was obtained as a colorless oil: 1.09 g (45%); ¹H NMR ($CDCl_3$) δ 1.43 (s, 9 H), 1.6-1.9 (m, 2 H), 2.00 (s, 2 H), 3.12 (q, 2 H), 3.71 (t, 2 H), 4.82 (s, 2 H), 5.12 (s, 1 H, NH), 7.45 (s, 5 H); mass spectrum (CI with Ar), m/e 323 (M + 1). In addition, a small amount of the hydroximate 12a was obtained from the earlier column fraction (about 11%). Compound 12a was crystallized from chloroform-/hexane: mp 86.5-88.5 °C; ¹H NMR (CDCl₃) δ 1.43 (s, 9 H), 1.92 (s, 3 H), 1.7–2.0 (m, 2 H), 3.30 (q, 2 H), 4.10 (t, 2 H), 4.98 (s, 2 H), 5.40 (s, 1 H, NH), 7.40 (s, 5 H).

1-[(tert-Butoxycarbonyl)amino]-5-[acetyl(benzyloxy)amino]pentane (4b). 5-[(tert-Butoxycarbonyl)amino]-1-pentyl bromide (11b; 1.33 g, 5 mmol), O-benzyl acetohydroxamate (6; 1.65 g, 10 mmol), KI (0.17 g, 1 mmol), and anhydrous K_2CO_3 (3.68 g, 40 mmol) were placed in dry acetone (25 mL) and refluxed for 24 h. The reaction mixture was filtered, evaporated, dissolved in ether, and washed with 0.5 N NaOH to remove excess 6. After the mixture was dried and the solvent evaporated, the residue was chromatographed on silica gel (2 \times 50 cm), eluting with ethyl acetate/hexane (35:65) to yield the product **4b**: colorless oil; 1.08 g (62%); ¹H NMR (CDCl₃) δ 1.1–1.8 (m, 15 H), 2.07 (s, 3 H), 3.08 (q, 2 H), 3.72 (t, 2 H) 4.80 (s, 2 H), 4.90 (s, 1 H, NH), 7.41 (s, 5 H). Anal. Calcd for C₁₉H₃₀N₂O₄: C, 65.11; H, 8.63; N, 7.99. Found: C, 64.99; H, 8.85; N, 7.96.

General Procedure for Alkylation of Hydroxamates 7-9 with Alcohol 10a Mediated by PPh₃/DEAD or Diisopropyl Azodicarboxylate (DIPAD). The alcohol 10a as an approximately 0.1 M solution in dry THF was treated with the hydroxamate (7, 8, or 9, 1.2 equiv) and PPh₃ (1.3 equiv). To this solution was added DEAD or DIPAD (1.3 equiv in a small amount of dry THF) dropwise over about 0.5 h with stirring at room temperature. The reactions were generally complete within an additional hour at room temperature, but if TLC indicated the presence of the starting alcohol, the reaction was left overnight. Products were characterized after isolation by spectral and elemental analysis when possible. Chromatography on silica gel was the standard method of isolation when 7 was alkylated with 10a by the general procedure. 13a was isolated in 78% yield by chromatography, eluting with ethyl acetate/hexane (20:80). The product was a colorless oil: ¹H NMR (CDCl₃) δ 1.46 (s, 9 H), 1.6-1.9 (m, 2 H), 2.01 (s, 3 H), 3.11 (q, 2 H), 3.53 (t, 2 H), 4.86 (s, 3 H, including NH), 5.23 (s, 2 H), 7.42 (2 s, 10 H). Anal. Calcd for C₂₀H₃₀N₂O₅: C, 66.65; H, 7.30; N, 6.76. Found: C, 66.34; H, 7.37; N, 6.64.

When compound 9 was alkylated with 10a by the general procedure, 13b was isolated in 89% yield by chromatography, eluting with CH_2Cl_2 . The product was a colorless oil: ¹H NMR $(CDCl_3) \delta$ 1.46 (s, 9 H), 1.6–1.9 (m, 2 H), 3.13 (q, 2 H), 3.57 (t, 2 H), 4.75 (s, 1 H, NH), 4.90 (s, 2 H), 5.30 (s, 2 H), 7.43 (s, 5 H), 7.55 (d, 2 H), 8.30 (d, 2 H).

When compound 8 was alkylated with 10a by the general procedure, 21 was isolated in 41% yield by chromatography, eluting with ethyl acetate/hexane (20:80). The product was a colorless oil: ¹H NMR (CDCl₃) δ 1.43 (s, 9 H), 1.50 (s, 9 H), 1.60–1.80 (m, 2 H), 3.10 (q, 2 H), 3.47 (t, 2 H), 4.83 (s, 3 H, including NH), 7.40 (s, 5 H). Anal. Calcd for C₂₀H₃₂N₂O₅: C, 63.13; H, 8.48; N, 7.36. Found: C, 62.99; H, 8.59; N, 7.16.

Compound 21 can be prepared from 11a (1 equiv) and 8 (1.1 equiv) by treatment with NaH (1.3 equiv) in DMF at 100 °C for 3 h. The reaction mixture was taken into ethyl acetate and washed with H_2O five times and once with brine. After the mixture was dried and the solvent evaporated, the residue was chromatographed on silica gel, eluting with ethyl acetate/hexane (20:80). The product 21 was isolated in 70% yield.

General Procedure for Exchange of the Cbz Group with an Acetyl Group. Compound 13, as an approximately 0.1 M solution in ethyl acetate, was treated with Ac₂O (2.5 equiv) and 5% Pd on carbon (15-20%) of the weight of 13) and stirred under 1 atm of H_2 at 0 °C for 3 h. The reaction mixture was filtered and evaporated, and the residue was taken up in ether. This was washed twice with 0.5 M NaOH and once with H_2O and brine. After the mixture was dried (MgSO₄) and the solvent evaporated, the crude product was chromatographed on silica gel, eluting with ethyl acetate/hexane (35:65). When 13a was converted to 4a by the general procedure, 4a was isolated in 65% yield (optimum). When 13b was converted to 4a by the general procedure, 4a was isolated in 55% yield (optimum). This product had identical spectral and TLC properties when compared with those for the previous preparation of 4a. Compound 14 can be prepared under the same conditions with a longer reaction time (5 h). If the reaction mixture was subsequently treated with CH₃OH and a catalytic amount of NH_3 for an additional 5 h, compound 15 was isolated as an oil in quantitative yield: ¹H NMR (acetone- d_{β}) δ 1.40 (s, 9 H), 1.6-1.9 (m, 2 H), 2.04 (s, 3 H), 3.08 (q, 2 H), 3.63 (t, 2 H), 6.0 (s, 1 H, OH).

1-Amino-5-[acetyl(benzyloxy)amino]pentane (16b). 1-[(tert-Butoxycarbonyl)amino]-5-[acetyl(benzyloxy)amino]pentane (4b; 0.526 g, 1.5 mmol) was stirred with CF_3CO_2H (2.0 mL) for 15 min at room temperature. Excess CF_3CO_2H was removed by rotary evaporation. The residue was partitioned between $CHCl_3$ (30 mL) and 10% Na₂CO₃. The chloroform layer containing free amine 16b was dried (K_2CO_3) and concentrated to give 0.345 g (92%) of 16b as an oil: ¹H NMR ($CDCl_3$) δ 1.1–1.8 (m, 6 H), 2.07 (s, 3 H), 2.63 (t, 2 H), 3.63 (t, 2 H), 4.80 (s, 1 H), 7.42 (s, 5 H).

1-Amino-3-[acetyl(benzyloxy)amino]propane Hydrochloride Salt (16a). 1-[(tert-Butoxycarbonyl)amino]-3-[acetyl(benzyloxy)amino] propane (4a; 1.8 g, 5.58 mmol) was dissolved in anhydrous ether (30 mL) at 0 °C. A stream of HCl gas was passed through this solution for 15-30 min. The solvent was evaporated to provide 16a in 97% yield as a white solid (very hygroscopic): mp 69-72 °C; ¹H NMR (D₂O) δ 1.7-2.1 (m, 2 H), 2.0 (s, 3 H), 2.92 (t, 2 H), 3.73 (t, 2 H), 4.83 (s, 2 H), 7.43 (s, 5 H). Anal. Calcd for C₁₂H₁₉N₂O₂Cl: C, 55.70; H, 7.40; N, 10.83; Cl, 13.70. Found: C, 55.24; H, 7.19; N, 10.53; Cl, 13.89.

1-(Acetylamino)-3-[(benzyloxy)amino]propane (18). Compound 4a (0.322 g, 1 mmol) was stirred with CF₃CO₂H (2 mL) for 15 min at room temperature. Excess CF₃CO₂H was removed by rotary evaporation. The residue was partitioned between CHCl₃ (30 mL) and 10% Na₂CO₃. After the mixture was dried (MgSO₄) and the solvent evaporated, the residue contained more than two products. It was chromatographed on silical gel, eluting with CH₂Cl₂/*i*-PrOH (90:10). Compound 18 was isolated in 52% yield as a colorless oil: ¹H NMR (CDCl₃) δ 1.5–1.8 (m, 2 H), δ 1.89 (s, 3 H), 2.92 (t, 2 H), 3.25 (q, 2 H), 4.70 (s, 2 H), 5.4 (s, 1 H, OH), 6.37 (s, 1 H, NH). Compound 18 was also isolated from the coupling reaction (see Scheme VII) and had the same spectral and TLC properties.

1-Amino-3-[(benzyloxy)amino]propane p-Toluenesulfonic Acid Double Salt (19). (a) Preparation from 4a. Compound 4a (0.322 g, 1 mmol) was dissolved in dioxane (10 mL), and TsOH (0.379 g, 2.2 mmol) was dissolved in dioxane (10 mL) separately. Both solutions were combined, and H_2O (1.5 equiv) was added. The reaction mixture was allowed to stand for 3 days. Ether was added to the reaction mixture, and 19 crystallized out: 72% yield (0.384 g); mp 150–152 °C; ¹H NMR (D₂O) δ 1.9–2.3 (m, 2 H), 2.40 (s, 6 H), 3.15 (t, 2 H), 3.50 (t, 2 H), 5.21 (s, 2 H), 7.57 (d, 4 H), 7.71 (s, 5 H), 7.95 (d, 4 H). Anal. Calcd for $C_{24}H_{32}N_2O_7S_2\cdot0.5H_2O$: C, 54.02; H, 6.23, N, 5.25. Found: C, 53.93; H, 6.09; N, 5.34.

(b) Preparation from 18. Compound 18 (0.11 g, 0.5 mmol) and TsOH (0.189 g, 1.1 mmol) were dissolved in dioxane/ H_2O (7:3, 10 mL). The reaction mixture was allowed to stand for 7 days. After evaporation of the solvent, the residue was recrystallized from methanol/ether to provide 19 in 75% yield (0.20 g).

(c) Preparation from 21. Compound 21 (1.18 g, 3.1 mmol) and TsOH (1.17 g, 6.82 mmol) were dissolved in dioxane/ H_2O (7:3, 50 mL). The solvent was evaporated with rotary evaporater at 50 °C. After removal of solvent, a white precipitate was formed which was recrystallized from methanol/ether to yield 1.35 g (81.7%) of 19. All three preparations provided 19 with the same melting point and ¹H NMR spectrum.

Coupling Reaction of Anhydromethylenecitryl Chloride 24 with Amine Hydrochloride 16a. Amine hydrochloride 16a (0.382 g, 1.47 mmol) and anhydromethylenecitryl chloride 24^{17,18} (0.177 g, 0.73 mmol) were dissolved in CHCl₃ (50 mL). Et₃N (0.297 g, 2.94 mmol) in CHCl₃ (25 mL) was added dropwise with stirring at 0 °C. After the addition was complete, the reaction mixture was allowed to warm to room temperature and stirred an additional 1 h. Then the solution was washed once with 0.5 M citric acid (aqueous), once with H₂O, 0.5 M NaHCO₃, and finally with brine. Then it was dried (MgSO₄) and evaporated to give a yellow oil which was contaminated by other side products (¹H NMR showed at least 90% desired product 25). The oil was chromatographed on silica gel (2 × 50 cm), eluting with CH₂Cl₂/*i*-PrOH (90:10). The separation was not successful due to the decomposition of 25.

2-Hydroxy-1,2,3-propanetricarboxylic Acid 2-Benzyl Ester (27a). Anhydromethylenecitric acid 23^{18} (2.04 g, 0.10 mmol) and Et₃N (3.06 mL, 22 mmol) were dissolved in CHCl₃ (30 mL). Benzyl alcohol (3.24 g, 30 mmol) was added, and the reaction mixture was refluxed for 3 days. The reaction mixture was added to ethyl acetate (100 mL) and extracted twice with 0.5 M NaHCO₃ (100 mL). The aqueous layer was adjusted to pH 2 with concentrated HCl, and then it was reextracted with ethyl acetate (200 mL) three times. After the mixture was dried (MgSO₄) and the solvent evaporated the residue was recrystallized from ethyl acetate (or ethyl acetate/hexane) to yield 27a: 1.61 g (57%); colorless crystals; mp 128–129 °C; ¹H NMR (CDCl₃) δ 2.88 (d, 4 H), 5.20 (s, 2 H), 7.40 (s, 5 H), 7.87 (br s, 3 H, CO₂H and OH). 2-Hydroxy-1,2,3-propanetricarboxylic Acid 2-Methyl Ester (27b). Anhydromethylene citric acid 23 (0.51 g, 2.5 mmol) and Et₃N (0.8 mL, 5.5 mmol) were dissolved in methanol (20 mL) and refluxed for 12 h. The reaction mixture was passed through a column of Dowex 50-X8 (H⁺); 50-mL bed) and washed through with a further 60 mL of H₂O. The solution was evaporated at reduced pressure. The residue was recrystallized from methanol/ether to yield 27b: 0.38 g (74%); mp 166–168 °C (lit.²³ mp 167 °C); ¹H NMR (acetone-d₆) δ 2.87 (d, 4 H), 3.75 (s, 3 H).

2-Hydroxy-1,2,3-Propanetricarboxylic Acid 2-Isopropyl Ester (27c). Anhydromethylene citric acid 23 (0.51 g, 2.5 mmol) and Et₃N (0.8 mL, 5.5 mmol) were dissolved in *i*-PrOH (15 mL) and refluxed for 21 h. The reaction mixture was passed through a column of Dowex 50-X8 (H⁺; 50-mL bed) and washed through with a further 60 mL of H₂O. The solution was evaporated at reduced pressure. The residue was recrystallized from acetone-/hexane to yield 27c: 0.44 g (75%); colorless crystals; mp 126-128 °C; ¹H NMR (acetone- d_6) δ 1.27 (d, 6 H), 2.88 (d, 4 H), 4.9-5.2 (m, 1 H).

General Procedure for the Preparation of Citric Acid Triester Derivatives 28. The diacid 27 and p-nitrophenol (1.3 equiv) were dissolved in dry acetonitrile to give a 0.1 M solution. The solution was cooled in an ice bath, and DCC (1.3 equiv in a small amount of dry acetonitrile) was added all at once. The solution was stirred 0.5 h at 0 °C, warmed to room temperature, and stirred an additional 5 h. After the dicyclohexyl urea was filtered off, the residue was chromatographed on silica gel eluting with CH₂Cl₂. The solution containing product was washed five times with saturated NaHCO₃ and brine and dried (MgSO₄). The solvent was evaporated, and the residue was recrystallized from ethyl acetate/hexane to give 28. 28a: 41% yield; mp 142.5-143.5 °C; ¹H NMR (CDCl₃) δ 3.32 (d, 4 H), 5.28 (s, 2 H), 7.32 (d, 4 H), 7.40 (s, 5 H), 8.32 (d, 4 H). 28b: 40% yield; mp 124-126 °C; ¹H NMR (CDCl₃) δ 3.22 (s, 4 H), 3.90 (s, 3 H), 4.10 (s, 1 H), 7.32 (d, 4 H), 8.32 (d, 4 H). 28c: 42% yield; mp 138-140 °C; ¹H NMR (CDCl₃) & 1.30 (d, 6 H), 3.21 (s, 4 H), 4.13 (s, 1 H, OH), 5.0-5.3 (m, 1 H), 7.30 (d, 4 H), 8.30 (d, 4 H).

Coupling Reaction of Amine Hydrochloride 16a with Activated Ester 28a. Activated ester 28a (100 mg, 0.194 mmol) and amine hydrochloride 16a (110 mg, 0.444 mmol) were suspended in acetonitrile (20 mL), and Et₃N (0.116 mL, 0.832 mmol) in acetonitrile (5 mL) was added dropwise. The resulting solution was allowed to stir for 3.5 h at room temperature. After the solvent was evaporated, preparative TLC (silica, 90% ethyl acetate-10% *i*-PrOH) was performed. Most of the product obtained was the acyl-transfer product 18. Very small amounts (<10%) of 29 and 30 were isolated. 29a: ¹H NMR (CDCl₃) δ 1.6-1.9 (m, 4 H) 2.07 (s, 6 H), 2.72 (s, 4 H), 3.20 (q, 4 H), 3.68 (t, 4 H), 4.82 (s, 4 H), 5.20 (s, 2 H), 7.10 (t, 2 H, NH), 7.36 (s, 5 H), 7.43 (s, 10 H). 30a: ¹H NMR (CDCl₃) δ 1.6-1.9 (m, 2 H), 2.74 (d, 2 H), 3.0-3.3 (m, 4 H), 3.70 (t, 2 H), 4.83 (s, 2 H), 5.24 (s, 2 H), 7.17 (d, 2 H), 7.38 (d, 10 H), 8.25 (d, 2 H).

Coupling Reaction of Amine p-Toluenesulfonic Acid Double Salt 19 with Activated Ester 28. Activated ester 28 and 19 (2.4 equiv) were suspended in acetonitrile (~ 0.1 M solution). Et_3N (6.8 equiv) in acetonitrile was added dropwise. The solution was allowed to stir 1 h at room temperature. It was then taken into ethyl acetate and washed with 10% Na₂CO₃ several times and once with H₂O and brine. After the mixture was dried $(MgSO_4)$ and the solvent evaporated, the residue was dissolved in acetonitrile, and acetic anhydride (3 equiv) was added. This solution was allowed to stir 2-3 h at room temperature. After evaporation the residue was chromatographed on silica gel, eluting with ethyl acetate/i-PrOH (90:10). When 19 was coupled with 28a, two main products 26 and 29a, were observed (85% yield, 26/29a ratio of 5:7) 26: oil; ¹H NMR (CDCl₃) & 1.6-1.9 (m, 4 H), 2.05 (d, 6 H), 2.70 (s, 2 H), 2.82 (s, 2 H), 3.20 (t, 2 H), 3.35-3.75 (m, 6 H), 4.80 (d, 4 H), 7.13 (t, 1 H, NH), 7.42 (s, 10 H). When 19 was coupled with 28b, two products, 26 and 29b, were obtained (82% yield, 26/29b ratio of 3:2 by ¹H NMR). When 19 was coupled with 28c only one product, 29c, was obtained in 75% yield without chromatography as a colorless oil. 29c: ¹H NMR (CDCl₃) δ 1.24 (d, 6 H), 1.5–1.85 (m, 4 H), 2.07 (s, 6 H), 2.67 (s, 4 H), 3.20

(q, 4 H) 3.70 (t, 4 H), 4.81 (s, 4 H), 4.97–5.25 (m, 1 H), 7.0 (t, 2 H, NH), 7.42 (s, 10 H); IR (neat) 1730, 1640 cm⁻¹.

Tribenzylarthrobactin 31. Activated ester 28a (0.27 g, 0.46 mmole) was dissolved in acetonitrile (30 mL). Amine 16b (0.27 g, 1.07 mmol) in acetonitrile was added, followed by addition of Et_3N (0.128 mL, 0.92 mmol). The reaction mixture was allowed to stir 2.5 h at room temperature. The reaction mixture was taken up in ethyl acetate (70 mL) and washed with phosphate buffer (pH 8.5, 1 M) several times and then once with water and brine. After the mixture was dried (MgSO₄) and the solvent evaporated, 0.329 g (95.6%) of pure 31 was isolated as an oil: ¹H NMR (CDCl₃) δ 1.2–1.8 (m, 12 H), 2.07 (s, 6 H), 2.67 (d, 4 H), 3.15 (q, 4 H), 3.63 (t, 4 H), 4.72 (s, 4 H), 5.18 (s, 2 H), 7.0 (t, 2 H, NH), 7.36 (s, 5 H), 7.44 (s, 10 H); IR (neat) 1730, 1640 cm⁻¹.

Methylschizokinen 32. Compound 29a (18 mg, 0.026 mmol) was dissolved in methanol (15 mL) and treated with 5% Pd on carbon (7 mg). The mixture was stirred at room temperature under 1 atm of H₂ for 7 h, filtered, and evaporated to give 32 as a hydroscopic, slightly off-white powder: 9.6 mg (85%); ¹H NMR (D₂O) δ 1.7-1.9 (m, 4 H), 2.02 (s, 6 H), 2.64 (s, 4 H), 3.17 (t, 4 H), 3.45-3.75 (m, 7 H); IR (CHCl₃) 1730, 1630 cm⁻¹.

Schizokinen A (33). Compound 26 (90 mg, 0.154 mmol) was dissolved in THF/water (7:3, 15 mL) and treated with 5% Pd on carbon (90 mg). The reaction mixture was stirred at room temperature under 1 atm of H₂ for 3 h and then filtered and evaporated to give pure 33 (48 mg, 77%) as white hydroscopic powder: ¹H NMR (D₂O) δ 1.5–1.9 (m, 4 H), 2.03 (s, 6 H), 2.6–3.2 (m, 6 H), 3.4–3.8 (m, 6 H); IR (CHCl₃) 1780 (weak), 1710, 1625 cm⁻¹; paper chromatography R_f 0.73 [butanol/water/acetic acid, 60:25:15 v/v (lit.¹⁰ R_f 0.74)]; mass spectrum (FD), m/e 402 (M⁺), 403 (M + 1), 359 [M – (CH₃C=O⁺)], 445 [M + (CH₃C=O⁺)].

Schizokinen (1). Compound 29c (55 mg, 0.086 mmol) was dissolved in THF (5 mL) and treated with NaOH (0.525 N, 175 μ L). The reaction mixture was stirred for 2 h (reaction was followed by TLC). Without isolation and characterization of 34a, the reaction mixture was treated with 5% Pd on carbon (50 mg) under 1 atm of H₂ for 3 h at room temperature. The reaction mixture was filtered and passed through a column of Dowex 50-X8 (H⁺; 20-mL bed). After a further 30 mL of H₂O was eluted, the solution was evaporated to give 31 mg (85%) of 1 as a powder. The 90-MHz ¹H NMR (D₂O)¹⁰ and the IR spectra²⁴ were identical with those depicted in the literature for natural schizokinen. Paper chromatography (butanol/H₂O/acetic acid, 60:25:15, and *i*-PrOH/H₂O, 7:3) gave the same R_f values (0.61, 0.66) as reported for the natural substance (0.60, 0.64):¹⁰ mass spectrum (FD), m/e 402 (M - H₂O), 403 (M + 1 - H₂O), 445 [M - H₂O + (CH₃C=O⁺)].

Arthrobactin (2). Compound 31 (120 mg, 0.161 mmol) was dissolved in THF (7 mL) and treated with 1 M NaOH (20 μ L). The reaction mixture was stirred at room temperature for 2 h. Without isolation of **34b** the reaction mixture was treated with 5% Pd on carbon (120 mg) under 1 atm of H₂ for 3 h at room temperature. The reaction mixture was filtered and passed through a column of Dowex 50-X8 (H⁺; 30-mL bed). After elution with 30 mL more of H₂O, the solution was evaporated at reduced pressure to give 2 as a hygroscopic, slightly off-white powder: 52.5 mg (68.6%). The 90-MHz ¹H NMR (D₂O) and IR spectra were in good agreement with those reported for the natural material.¹¹ In butanol/H₂O/acetic acid (60:25:15) and in *i*-PrOH/H₂O (7:3) they gave R_i values of 0.81 and 0.81, respectively: mass spectrum (FD), m/e 459 (M + 1 - H₂O), 458 (M - H₂O).

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New Construction of a Steroidal Ring System. Stereoselective Synthesis of (\pm) -Androstane-2.17-dione

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Intramolecular Diels-Alder reaction of the triene 3 afforded the tricyclic olefin 4 which, after deprotection of the ketal groups, was treated with ethylaluminum dichloride to give (\pm) -androstane-2,17-dione (6) stereoselectively.

In a recent development toward the synthesis of steroids, the intramolecular cycloaddition reaction has played an important role because of its effective regio- and stereoselectivity. Among the numerous reports using it as a key step, much attention has been focused on the synthesis of A-aromatic steroids^{1,2} such as estrone and estradiol and on the stereocontrolled construction of trans-hydrindan ring systems.³ We now report a novel stereoselective synthesis of a nonaromatic steroid employing an intramolecular Diels-Alder reaction and a subsequent Lewis acid catalyzed ring-closure reaction as key steps.

Results and Discussion

For the purpose of accomplishment of our synthetic strategy, illustrated in Scheme I, the dienophile 1 was prepared from Hagemann's ester according to the method reported by us,⁴ whereas the diene 2 was synthesized as follows (Scheme II).

Ethyl 2,5,5-trimethyl-1,3-dioxane-2-acetate (7),⁵ on treatment with lithium aluminum hydride (3 equiv) in tetrahydrofuran at ambient temperature afforded the alcohol 8, which was then converted to the iodide 10 via the tosylate 9 by tosylation of 8 with p-toluenesulfonyl chloride (1.5 equiv) and pyridine (2 equiv) in methylene chloride

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Scheme I



 $\frac{4}{2}$ R¹ = O(CH₂)₂O, R² = OCH₂C(CH₃)₂CH₂O $5 R^1 = R^2 = 0$



and subsequent treatment of 9 with sodium iodide (5 equiv) in acetone (65% yield from 7). Regioselective alkylation of crotonaldehyde with the iodide 10 has been carried out by using the Schiff base 11 and lithium diisopropylamide (1.1 equiv) under the conditions reported by Schlessinger⁶ to afford the α -alkylated product 12 in 45% yield. Wittig methylenation⁷ of 12 with triphenylmethylphosphonium bromide (2.5 equiv) and n-butyl-

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